

The role of exercise on the innate immunity of the elderly

Lucia Malaguarnera · Erika Cristaldi · Maria Vinci ·
Mariano Malaguarnera

Received: 22 August 2007 / Accepted: 8 October 2007 / Published online: 13 November 2007
© EGREPA 2007

Abstract The increased life span in human population has shown that some diseases, as infections, cancer and autoimmune phenomena, occur more frequently in the elderly than in the younger. We describe the ageing process involving the innate immune system and the improvement given by moderate physical activity. In addition, we discuss the altered neutrophil granulocytes function, the role of macrophages and natural killer cells, besides the influence of cytokines and secretory IgA. The acquired information help us to explain how these changes could favor the onset of diseases in the elderly and how they may boost their immune function.

Keywords Immunosenescence · Neutrophil granulocytes · SIgA · Moderate exercise · Cytokines

Introduction

Ageing may be considered as a slow and progressive preparation of the organism for a morphological and functional involution, which takes part in the biological cycle [39]. Many changes, involving systems, apparatuses and organs, occur during this continuous process. The immune function is also involved, thereby contributing to the increased susceptibility to infections, cancer and autoimmunity diseases [31].

The immune response consists of two interactive components, the innate (natural) and the acquired (adaptive) response, both of them providing the primary defence against pathogens. Many evidences have suggested that acquired immunity is primarily affected by changes in T-cell function, with B-cell function relatively locked. The modifications of the innate immune system include altered phagocytic capacity, decreased generation of radicals and dysfunctional cytokines production [37].

In the elderly, the proinflammatory phenotype [in particular, the elevation of plasma levels of tumour necrosis factor- α (TNF- α) and interleukin (IL)-6] is associated with an increased morbidity and mortality. In contrast, the anti-inflammatory phenotype, especially IL-10 levels, may be associated with longevity [8].

In the elderly often, a progressive increase in the natural killer (NK) cell phenotype has been noticed. In contrast, in the centenarians, the increase in the NK cells correlated with a good functionality that did not result in meaningful decrease in comparison to the young people [20].

Physical activity and immune function

Inflammatory state, cancer and autoimmune diseases are very common in the elderly. We know the role of immune function in the onset of these illnesses. A compromised function of innate immunity leads to higher susceptibility to upper respiratory tract infections (URTI) such as cold or influenza [56]. Growing evidence revealed that regular and moderate physical activity could promote increase in the innate immune functions (Table 1). It has been demonstrated that the incidence and mortality rates for certain types of cancer, particularly tumours of the colon and the female reproductive tract, are lower among active subjects [57].

Exercise programs also appear to have a beneficial influence on clinical course, at least in the early stages of the disease. The

L. Malaguarnera
Department of Biomedical Sciences, University of Catania,
Catania, Italy

E. Cristaldi · M. Vinci · M. Malaguarnera (✉)
Department of Senescence, Urological and Neurological Sciences,
University of Catania,
Azienda Ospedaliera Cannizzaro, via Messina 829, Cannizzaro,
95124 Catania, Italy
e-mail: malaguar@unict.it

Table 1 Variations of innate immunity in the elderly through physical activity

	Effect of ageing	Effect of moderate exercise
NK cell numbers	Increased	Increased
NKCA	Decreased	Increased or unchanged
Neutrophil numbers	Unchanged	Increased
Neutrophil function	Decreased	Increased
Monocyte/macrophage function	Decreased	Increased
Pro-inflammatory cytokines	Increased	Low decreased
Anti-inflammatory cytokines	Decreased	Low increased
SIgA secretion rate	Decreased	Increased

NK Natural killer, NKCA natural killer cell activity, SIgA secretory IgA

role of the immune system may be limited, however, depending on the sensitivity of the specific tumour to cytolysis, the stage of cancer, the type of exercise program and many other complex factors [43].

Regular moderate physical activity may reduce URTI symptoms; in contrast, heavy acute or chronic exercise may increase the risk of upper respiratory tract infection [30]. Many reports suggest that infection severity, relapse and myocarditis may result when patients exercise vigorously [21]. In response to acute exercise, it has been observed that a rapid interchange of immune cells between peripheral lymphoid tissues and the circulation occurs [45]. The response depends on the intensity, duration and mode of exercise, changes in body temperature, blood flow, hydration status and body position concentrations of hormones and cytokines. The cells showing more responsiveness to the effects of acute exercise, both in terms of numbers and function, are NK cells, neutrophils and macrophages [60]. Instead, helper T-cell counts and other immune measures are not enhanced significantly. In response to long-term exercise training, NK-cell activity shows a significant elevation. Some evidence indicates that neutrophil function is suppressed during periods of heavy training. It has been reported that acute exercise bouts of moderate duration (<60 min) and intensity [<60% oxygen uptake (VO₂max)] are associated with fewer perturbations and less stress to the immune system than are prolonged, high-intensity sessions [62].

Neutrophil granulocytes

The most frequent infectious diseases in elderly people affect the upper respiratory tract, increasing the risk of mortality in the population with higher susceptibility because of the lowered immune defence.

The primary barrier to bacteria and fungi attack is represented by polymorphonuclear cells. Various studies,

comparing the old and the young population, have shown that neutrophil numbers in blood and in bone marrow are still the same [4]. Moreover, chemotaxis and vascular adhesion did not show changes in old adults [3]. Therefore, the compromised function of neutrophil granulocytes could be due to altered phagocytosis and decreased bactericidal capacity, which deals with degranulation and superoxide generation [22]. An explanation for this inability has been given by studying the expression of such membrane markers, as CD16, CD32, CD64 and CD11b. In detail, Butcher et al. [6] have identified an Fc-gamma receptor, called CD16, which declines with age. They noticed neutrophilia during bacterial infection but lowered level of CD16. It has been suggested that the origin of alteration arises in the bone marrow [6]. These findings could contribute to give more information about the reduced production of superoxide anion in response to Gram-positive bacteria, as *Staphylococcus aureus*. Presumably, these types of microbes need Fc-receptor and complement bond.

Escherichia coli, rather, depends on CD14 binding, likely unaffected by ageing; thus the elderly are more susceptible to *S. aureus* and other Gram-positive infections.

In this last mechanism, the presence of ion calcium is not required; thereby, the reactive oxygen intermediates do not undergo any changes.

The calcium intracellular concentration influences the phagocytosis and ability of killing microbes. The increase in calcium levels leads to activation of neutrophils granulocytes; thus, Wenisch et al. [67] hypothesized that inability of neutrophils may be due to elevated resting ion calcium levels; moreover, with ageing, the extrusion of calcium is reduced after stimulation with f-Met-Leu-Phe [23].

Indeed, the same group found that a decreased transport of esoses, fuel for the neutrophils, might be responsible for reduced functionality of these cells [67].

Long-lasting and severe URTI in elderly could not be due to the reduced numbers of neutrophils but to lowered expression of some membrane markers, involving the transmembrane signalling and the production of reactive species, which intervene in the killing of microbes.

Exercise and neutrophils

Relatively few studies have been carried out on various athletes to study the role of neutrophils during exercise. Exercise promotes the release of neutrophils into the circulation, depending on the increase in both catecholamine and cortisol levels. Neutrophils mobilised after exercise have an enhanced capacity to generate some forms of reactive oxygen species (ROS) when stimulated in vitro [50].

ROS generated during intense exercise may lead to DNA damage in leukocytes, but it is unknown if this damage is the result of neutrophil activation.

Following intensive and prolonged exercise training, the function of neutrophils included seems to be impaired [46].

In elite swimmers undergoing strong training, the oxidative activity of neutrophils is markedly lower than controls [55]. Instead, elite female rowers did not show alterations in granulocyte and monocyte phagocytosis and oxidative burst activity before a World Championship [47]. Furthermore, after downhill running in well-trained runners, alterations were absent in degranulation and respiratory burst activity [52]. However, at present, variation in exercise-induced alterations in neutrophil function could be dependent on the differences in exercise protocols and training status.

Macrophages

The macrophages exert a considerable role in immune response and act through several mechanisms as by phagocytosis and elimination of microbes and by the release of mediators like interleukin 1 (IL-1) and TNF- α . Moreover, they play a fundamental role in repairing tissue damage.

With increasing age, the number of monocytes does not appear to change appreciably [38]. Increasing experimental evidence supports the premise that haematopoietic stem cells age and have a limited functional life span [25]. This could explain the hypocellularity in the macrophage precursors observed in the bone marrow of elderly people [48]. Early works suggested that macrophages produce similar levels of cytokines and that difference in function could be modulated through changes in T- and B-cell responses to such substances [40]. Some authors found increased production of IL-1, IL-6, IL-8 and TNF- α by human macrophages, while others reported a decreased synthesis [9, 17]. The decreased production of IL-6 and TNF- α by monocytes from aged subjects following lipopolysaccharide stimulation has been associated with the deficiencies in the activation of protein kinase C (PKC) α , PKC- β_1 , PKC- β_{11} , mitogen-activated protein kinase and deficient expression of c-FOS and c-Jun [12]. Additionally, Gon et al. reported a decreased release of granulocyte-macrophage colony-stimulating factor, granulocyte colony-stimulating factor and the chemokine macrophage inflammatory protein 1 α (MIP 1 α) by human macrophages. Overall, these contradictory results could be explained by the differences in experimental conditions, the methods used to measure the levels of cytokines and the health status of subjects [27].

A cause of macrophage ageing is the acquisition of defects in genomic DNA by a combination of both DNA damage and impaired DNA repair capacity and, consequently, the loss of several functional activities.

Interestingly, a recent study showing the effect of a higher production of TNF- α in splenic macrophages incubated in vitro with autologous serum suggested that the age-specific external milieu exerts an effect on macrophage activation

[26]. In fact, monocytes/macrophages are exposed to many agents, like hormones, cytokines and fatty acids, which can act on phenotype and function, responsible for the functional plasticity and adaptation to changing microenvironments. The circulating levels of these agents may change in the elderly and have effects on macrophages [63].

Physical activity and macrophages

Macrophages are involved in skeletal muscle repair through pro-inflammatory and alternative functions. It has been supposed that ageing alters the abundance and properties of skeletal muscle macrophages that will influence their functional response to acute resistance exercise. Pre-exercise, young muscle tended to possess a greater number of macrophages, whereas elderly muscle possessed higher levels of IL-1 beta, Interleukin 1 receptor antagonist (IL-1Ra) and IL-10. Post-exercise, total macrophages did not change in either group. The number of CD11b+ and CD163+ cells increased only in the young. Both subpopulations increased their activity post-exercise exclusively in the young. This finding suggested that ageing results in a defective regulation of muscle macrophage function, both at baseline and in response to resistance exercise, that may limit muscle hypertrophy in older adults [54]. In a recent study in murine model, it has been shown that acute restraint stress is associated with impaired function of macrophages. Whereas, moderate physical training attenuates the effects of acute stress by a mechanism that involves an increase in tolerance of macrophages [36].

Natural killer

The natural killer cells play a critical role in the innate immune response against infections and tumours. NK are cytotoxic cells that differ from cytotoxic T cells by their ability to lyse targets without the requirement of an antigen sensitisation. Early studies suggested no changes in the cytotoxic ability of NK cells with age [34], whereas recently, new evidences contradict these findings. The number of NK cells appears to increase with ageing, but NK activity decreases [16, 35]. The functional decline of the NK cells has been attributed to various factors such as decreased responses to interferon (IFN)- α , IL-2 and IL-12 [5] and increased responses to the negative modulating adenosine triphosphate (ATP) [33].

In some studies, Moccheggiani et al. [42] reported the remarkable role of neuroendocrine-immune pathway in immunosenescence. Hypothalamic, pituitary, gonadal and thyroid hormones' receptors are present on NK cells and on many other immune cells to stimulate cytokine production, thereby affecting adaptive immune responses. Additionally, hormone deficiency, a peculiarity of the elderly people, leads to impaired immune responses. Furthermore, the bioavail-

ability of the ion zinc is relevant, which is important to maintain the efficacy of the neuroendocrine-immune network in ageing [42].

In contrast to the general age-related decline in T and B reactivity, the NK cell system is highly active in the majority of healthy elderly (>80 years) [32]. High NK activity was especially demonstrated in the >80-year group, suggesting that the rate of increase in age-specific cancer incidence tends to be slow and in the same cancers may actually decline in incidence over the age of 80 [10].

Physical activity and NK

In the NK cells, the intensity of exercise influences both the number and the activity. A single bout of exercise for at least 1 h causes cell decrease and a decline of the function; thus, the capacity to lyse tumour target cells is inhibited [51].

The response to chronic stress of an intensive exercise, in athletes compared with non-athletes, leads to increased NK cell activity (NKCA) [51]. In the old adults, a single bout of moderate exercise does not have influence on NKCA. A training program gives beneficial effects on resting NKCA, followed by increase control of both viral infections and of malignant cell formation [66].

Flynn et al. [19] observed a transient increase in NK cell number immediately after exercise in elderly women who underwent 10 weeks of resistance training. Moreover, a comparative study examining the relationship between active in respect to inactive lifestyles and immunocompetence showed that the concentration of NK-cells (CD16+CD56+) significantly increased in the elderly exercisers, compared to that of the age-matched control subjects, or of the young group. The phagocytotic activity of neutrophils showed an age-associated decline but of lesser degree in the elderly exercisers than in the elderly controls. These results suggest that habitual and moderate training in later life is associated with a lesser age-related decline in certain aspects of circulating T-cell function and innate immunity [68].

Another study demonstrated that the natural killer cells response to a single exercise challenge is normal in older individuals, but immediately after exercise, the elderly subjects manifest less suppression of phytohemagglutinin-induced lymphocyte proliferation than younger individuals. In contrast, a strenuous exercise seems to induce a more sustained post-exercise suppression of cellular immunity in older individuals than in young subjects. A few cross-sectional comparisons of immune status between physically fit elderly individuals and young sedentary controls suggest that habitual physical activity may enhance NK cell activity [59]. More recently, elderly men, who were under training regularly for more than 16 years, exhibited a NK cell percentage remarkably greater than those of control group [7].

Overall, only moderate exercise is able to enhance oxidative burst activity, blood granulocytes and monocyte phagocytosis [49, 62].

Cytokines

In the elderly subjects, lymphocytic population mostly represented are CD28⁻CD8⁺ T lymphocytes. These cells are responsible for an increased production of type 1 cytokines, as IFN- γ and TNF- α , that own defence activity and, on the other side, support a chronic inflammatory status [69]. This status is called “inflamm-ageing” and is likely due to a chronic antigenic load. The latter leads to peculiar increase in inflammatory cytokines and of acute-phase protein production (Table 2).

The inflammatory cytokines are able to improve the expansion and the survival of CD4⁺ T cell effectors [11], thereby improving responses to vaccine. This action is due to the overcoming of the reduced transcription factor activation in aged CD4 cells, thus enhancing IL-2 production, which in turn leads to enhanced CD4 effector generation [29].

Many studies reported a decreased IL-2 and an increased IL-4 production, which is an anti-inflammatory cytokine belonging to type 2 cytokines pattern. IL-4 is mostly produced by activated CD4⁺ cells than the virgin cells. Likely, its increased levels need to offset elevated serum levels of TNF- α and IFN- γ . Furthermore, Alberti et al. [2] have noticed a decreased ratio of IFN- γ /IL-4 in the ageing, suggesting a shift towards an increased role of pattern 2 than pattern 1.

IL-6 is an inflammatory mediator and may be involved in the pathogenesis of numerous age-associated pathologies such as various lymphoproliferative disorders [64]. An increase in IL-6 serum levels appears to be necessary to stimulate the hypothalamus [65].

An interesting research conducted on IL-15 in ultra-longevity subjects showed that this interleukin stimulated the proliferation of memory T cells (CD45RO⁺), both CD4 and CD8, which were the most represented in the elderly. Furthermore, IL-15 led to NK CD56⁺ cell differentiations, which have a key role on defence against bacteria, fungi,

Table 2 Changes of cytokines in elderly

Cytokines	Changes
IL-2	Decreased
IL-2R	Decreased
IFN- γ	Increased
TNF- α	Increased
IL-6	Increased
IL-15	Increased
IL-4	Increased
IL-10	Increased

IL Interleukin; IFN Interferon; TNF tumour necrosis factor

viruses and protozoa [18]. The enhanced levels seen in the centenarians may explain how the old subjects may protect themselves from infections [24].

Physical activity and cytokines

Some studies carried out on runners and marathoners have demonstrated increased blood concentrations of cytokines, both inflammatory and anti-inflammatory type. In contrast, moderate exercise for no more than 1 h did not seem to have appreciable effects in cytokines levels [61]. The highest level of a cytokine is that of IL-6, which increases after running for 6 h, reaching threefold the baseline level [14], and immediately after 2.5 h of running, IL-6 increased 5.5-fold [44]; besides, a rise of 100-fold has been detected on marathoners [51].

After 2 h by the end of exercise, it has been observed that there are increased levels of IL-1Ra, that is an inhibitor of IL-1, which intervenes to limit the harmful effect of IL-1. The up-regulation of IL-1Ra is due to several cytokines, including IL-6.

IL-1Ra rises 1.5 h post-run, while IL-6 increases during the run [44].

The transforming growth factor TGF- β has shown no enhanced levels, rather than TNF- α , which is increased about threefold in strenuous exercise.

Very interesting is the inhibitor role of IL-6, released in response to exercise-induced muscle damage, on TNF- α [52]. This inhibition may prevent some negative effects, leading to a better lipid profile, elevated insulin sensitivity, a lower blood pressure and anti-inflammatory activity [52]. Mostly, this anti-inflammatory effect may be due to regularly performed exercise [53].

Although it is an unexplored pathway, it could be hypothesized that there is a beneficial effect of exercise on the anti-inflammatory state of elderly people, if they maintain the capacity to release IL-6 during exercise. Resistance exercise, performed by old adults, have shown a decrease in TNF- α levels [28].

Even the intensity of the exercise has some influences on cytokine expression. That has been shown by a Polish study, which examined elderly women undergoing moderate-intensity exercise. The results demonstrated that the percentage of lymphocytes expressing intracellular IL-2 was higher than sedentary control women, and it was the same for young women [13]. The intracellular IL-4-expressing lymphocytes have been decreased, thus contributing to control higher levels of memory cells than naïve cells [13].

Secretory immunoglobulin-A

The secretory immunoglobulin-A (SIgA) are prevalent in saliva, mother's milk, tears, intestinal and bronchoalveolar secretions and in other mucosal fluids. They play a fundamental

role against pathogens, which get in through the mucosas. The immunoglobulins bind bacteria and virus, avoiding adhesion and colonization of pathogens. Thus, they prevent infections of upper respiratory tract, as cold or influenza, which are very frequent in old adults. Therefore, decreased levels of SIgA contribute to higher susceptibility of elderly to this kind of infections. Studies carried out in elderly people reported lower SIgA secretion rates at rest both in women and in men [41, 15].

Physical activity and immunoglobulins

Furthermore, surface immune defence could get some beneficial effects by physical activity, performed at low-intensity; thus the SIgA secretion rate, reduced as a consequence of immunosenescence, may enhance.

The SIgA secretion rate depends on IgA concentration and saliva flow rate.

Increased value in SIgA secretion rate was given by an enhancement of SIgA concentration in old adults after an endurance exercise program for 12 months [1].

Sakamoto et al. [56], instead, found a rise of saliva flow rate after exercise; likely, the parasympathetic nervous system is activated after exercise.

The enhancement of saliva flow and SIgA secretion rate is just temporary after low-intensity exercise [56]; instead, another study reported high salivary IgA levels after 12 months of training, although the mechanism is unknown [1].

Recently, it has been demonstrated that a moderate physical activity, as 7,000 steps per day, may improve mucosal immune function in the elderly [58].

Conclusions

A large evidence suggests that the immune changes represent one of the most important phenomena for understanding the pathogenesis of the age-associated diseases. The innate immune system is affected by the ageing process.

Following these data, it may be considered extremely useful for elderly people to have a moderate exercise program to reverse physiological changes in immune function which occur with ageing, increasing their life quality.

References

1. Akimoto T, Kumai Y, Akama T, Hayashi E, Murakami H, Soma R, Kuno S, Kono I (2003) Effects of 12 months of exercise training on salivary secretory IgA levels in elderly subjects. *Br J Sports Med* 37:76–79
2. Alberti S, Cevenini E, Ostan R, Capri M, Salvioli S, Bucci L, Ginaldi L, De Martinis M, Franceschi C, Monti D (2006) Age-dependent modifications of type 1 and type 2 cytokines within virgin and memory CD4⁺T cells in humans. *Mech Ageing Dev* 12:560–66

3. Biasi D, Carletto A, Dell'Agnola C, Caramaschi P, Montesanti F, Zavateri G, Zeminian S, Bellavite P, Bambara LM (1996) Neutrophil migration, oxidative metabolism, and adhesion in elderly and young subjects. *Inflammation* 20:673–681
4. Born J, Uthgenannt D, Dodt C, Nunninghoff D, Ringvold E, Wagner T, Fehm HL (1995) Cytokine production and lymphocyte subpopulations in aged humans. An assessment during nocturnal sleep. *Mech Ageing Dev* 84:113–126
5. Burns EA, Leventhal EA (2000) Aging, immunity and cancer. *Cancer Control* 7:513–520
6. Butcher SK, Chahal H, Nayak L, Sinclair A, Henriquez NV, Sapey E, O'Mahony D, Lord JM (2001) Senescence in innate immune responses: reduced neutrophil phagocytic capacity and CD16 expression in elderly humans. *J Leukoc Biol* 70:881–886
7. Buyukyazi G (2004) Differences in the cellular and humoral immune system between sedentary and endurance-trained elderly males. *Science and Sports* 19:130–135
8. Caruso C, Lio D, Cavallone L, Franceschi C (2001) Aging, longevity, inflammation and cancer. *Ann N Y Acad Sci* 1028:1–13
9. Clark JA, Peterson TC (1994) Cytokine production and aging: overproduction of IL8 in elderly males in response to lipopolysaccharide. *Mech Ageing Dev* 77:127–139
10. Crawford DH (1984) Lymphocyte and macrophage function. *Br J Hosp Med* 32(112):114–118
11. Curtsinger JM, Schmidt CS, Mondino A, Lins DC, Kiedl RM, Jenkins Mk, Mescher MF (1999) Inflammatory cytokines provide a third signal for activation of naïve CD4⁺ and CD8⁺ T cells. *J Immunology* 162:3256
12. Delpedro AD, Barjavel MJ, Mamdouth Z, Faure S, Bakouche O (1998) Signal transduction in LPS-activated aged and young monocytes. *J Interferon Cytokine Res* 18:429–437
13. Drela N, Kozdron E, Szczypiorski P (2004) Moderate exercise may attenuate some aspects of immunosenescence. *BMC Geriatrics* 4:8
14. Drenth JP, Van Uum SHM, Van Deuren M, Pesman GJ, Van Der VenJongekrijg J, Van Der Meer JWM (1995) Endurance run increases circulating IL-6 and IL-1ra but downregulates ex vivo TNF-a and IL-1 β production. *J Appl Physiol* 79:1497–1503
15. Eliasson L, Birkhed D, Osterberg T, Carlén A (2006) Minor salivary gland secretion rates and immunoglobulin A in adults and the elderly. *Eur J Oral Sci* 114(6):494–499
16. Facchini A, Mariani E, Mariani AR (1987) Increased number of circulating Leu 11+ (CD16) large granular lymphocytes and decreased NK activity during human ageing. *Clin Exp Immunol* 68:340–347
17. Fagiolo U, Cossarizza A, Scala E, Fanales-Belasio E, Ortolani C, Cozzi E, Monti D, Franceschi C, Paganelli R (1993) Increased cytokine production in mononuclear cells of healthy elderly people. *Eur J Immunol* 23:2375–2378
18. Fehniger TA, Caligiuri MA (2001) Interleukin 15: biology and relevance to human disease. *Blood* 97(1):14–32
19. Flynn MG, Fahlman M, Braun WA, Lambert CP, Bouillon LE, Brolinson PG, Armstrong CW (1999) Effects of resistance training on selected indexes of immune function in elderly women. *J Appl Physiol* 86(6):1905–1913
20. Franceschi C, Motta L, Motta M, Malaguamera M, Capri M, Vasto S, Candore G, Caruso C; IMUSCE (2007) The extreme longevity: The state of the art in Italy. *Exp Gerontol*. Jul 1 (in press)
21. Friman G, Larsson E, Rolf C (1997) Interaction between infection and exercise with special reference to myocarditis and the increased frequency of sudden deaths among young Swedish orienteers 1979–92. *Scand J Infect Dis Suppl* 104:41–49
22. Fulop T, Komaromi I, Foris G, Worum I, Leovey A (1986) Age-dependent variations of intralysosomal release from human PMN leukocytes under various stimuli. *Immunobiol*. 171:302–310
23. Fulop T Jr, Hauck M, Worum I, Foris G, Leovey A (1987) Alterations of the fMLP-induced Ca²⁺ efflux from human monocytes with aging. *Immunol Lett* 14:283–286
24. Gangemi S, Basile G, Monti D, Merendino RA, Di Pasquale G, Bisignano U, Nicita-Mauro V, Franceschi C (2005) Age-related modifications in circulating IL-15 levels in humans. *Mediat Inflamm* 4:245–247
25. Geiger H, Van Zant G (2002) The aging of lympho-hematopoietic stem cells. *Nature Immunol* 3:329–333
26. Gomez CR, Boehmer ED, Kovacs EJ (2005) The aging innate immune system. *Curr Opin Immunol* 17:457–462
27. Gon Y, Hashimoto S, Koura T, Matsumoto K, Horie T (1996) Lower serum concentrations of cytokines in elderly patients with pneumonia and the impaired production of cytokines by peripheral blood monocytes in the elderly. *Clin Exp Immunol* 106:120–126
28. Greiwe JS, Cheng B, Rubin DC, Yarasheski KE, Semenkovich CF (2001) Resistance exercise decreases skeletal muscle tumor necrosis factor alpha in frail elderly humans. *FASEB J* 15:475–482
29. Haynes L, Eaton SM, Burns EM, Rincon M, Swain SL (2004) Inflammatory cytokines overcome age related defects in CD4⁺ T cell responses in vivo. *J Immunol* 172:5194–5199
30. Houston MS, Silverstein MD, Suman VJ (1997) Risk factors for 30-day mortality in elderly patients with lower respiratory tract infection. *Arch Intern Med* 157:2190–2195
31. Humme M, Paavilainen PM, Pertovaara M, Jylha M, Karhunen PJ, Hervonen A, Lehtimäki T (2005) Ig A levels are predictors of mortality in Finnish nonagenarians. *Mech Ageing Dev* 126(6–7):829–831
32. Krishnaraj R, Blandford G (1987) Age-associated alterations in human natural killer cells. 1. Increased activity as per conventional and kinetic analysis. *Clin Immunol Immunopathol* 45:268–285
33. Krishnaraj R (1992) Negative modulation of human NK cell activity by purinoceptors. 2. Age-associated, gender-specific partial loss of sensitivity to ATP. *Cell Immunol* 144:11–21
34. Kutza J, Kaye D, Murasko DM (1995) Basal natural killer cell activity of young versus elderly humans. *J Gerontol A Biol Sci Med Sci* 50:110–116
35. Kutza J, Murasko DM (1996) Age-associated decline in IL-2 and IL-12 induction of LAK cell activity of human PBMC samples. *Mech Ageing Dev* 90:209–222
36. Leandro CG, de Lima TM, Alba-Loureiro TC, do Nascimento E, Manhaes de Castro R, de Castro CM, Pithon-Curi TC, Curi R (2007) Stress-induced downregulation of macrophage phagocytic function is attenuated by exercise training in rats. *Neuroimmunomodulation* 14(1):4–7
37. Lio D, Malaguamera M, Maugeri D, Ferlito L, Bennati E, Scola L, Motta M Caruso C. (2007) Laboratory parameters in centenarians of Italian ancestry. *Exp Gerontol* Jul 1 (in press)
38. Malaguamera M, Laurino A, Di Mauro S, Motta M, Di Fazio I, Maugeri D (2000) The comorbidities of elderly oncologic patients. *Arch Gerontol Geriatr* 30:237–244
39. Malaguamera L, Ferlito L, Imbisi RM, Gulizia GS, Di Mauro S, Maugeri D, Malaguamera M, Messina A (2001) Immunosenescence: a review. *Arch Gerontol Geriatr* 31:1–14
40. McLachlan JA, Serkin CD, Morrey-Clark KM, Backouche O (1995) Immunological functions of aged human monocytes. *Pathobiology* 63:148–159
41. Miletic ID, Shiffman SS, Miletic VD, Sattely-Miller EA (1996) Salivary IgA secretion rate in young and elderly persons. *Physiol Behav* 60:243–248
42. Mocchegiani E, Giacconi R, Muti E, Muzzioli M, Cipriano C (2004) Plasticity of neuroendocrine-thymus interactions during ontogeny and aging: role of zinc. In: Straub Moccheggiani RE (ed) *Neuroimmunobiology, “the neuroendocrine immune network in aging”* vol.4. Elsevier, Amsterdam, pp 305–327
43. Murphy EA, Davis JM, Brown AS, Carmichael MD, Mayer EP, Ghaffar A (2004) Effects of moderate exercise and oat beta-glucan on lung tumor metastases and macrophage antitumor cytotoxicity. *J Appl Physiol* 97(3):955–959

44. Nehlsen-Cannarella SL, Fagoaga OR, Nieman DC, Henson DA, Butterworth DE, Schmitt RL, Bailey EM, Warren BJ, Utter A, Davis JM (1997) Carbohydrate and the cytokine response to 2.5 h of running. *J Appl Physiol* 82(5):1662–1667
45. Nieman DC, Nehlsen-Cannarella SL (1994) The immune response to exercise. *Semin Hematol* 31:166–179
46. Nieman DC, Hoffman-Goetz L (1996) In Hoffman-Goetz L (ed) Exercise and immune function. CRC, New York pp 143–162
47. Nieman DC, Nehlsen-Cannarella SL, Fagoaga OR, Henson DA, Shannon M, Hjertman JME, Schmitt RL, Bolton MR, Austin MD, Schilling BK, Thorpe R (2000) Immune function in female elite rowers and non-athletes. *Br J Sports Med* 34:181–187
48. Ogawa T, Kitagawa M, Hirokawa K (2000) Age-related changes of human bone marrow: a histometric estimation of proliferative cells, apoptotic cells, T-cells, B-cells and macrophages. *Mech Ageing Dev* 117:57–68
49. Ortega E (1994) Physiology and biochemistry: influence of exercise on phagocytosis. *Int J Sports Med* 15:S172–S178
50. Peake J, Suzuki K (2004) Neutrophil activation, antioxidant supplements and exercise-induced oxidative stress. *Exerc Immunol Rev* 10:129–141
51. Pederssen BK, Toft AD (2000) Effects of exercise on lymphocytes and cytokines. *Br J Sports Med* 34:246–251
52. Pedersen BK, Steensberg A, Fischer C, Keller C, Keller P, Plomgaard P, Febbraio M, Saltin B (2003) Searching for the exercise factor: is IL-6 a candidate? *J Muscle Res Cell Motil* 24:113–119
53. Pedersen BK, Febbraio MA (2005) Muscle derived interleukin-6: a possible link between skeletal muscle, adipose tissue, liver and brain. *Brain Behav Immun* 19(5):40
54. Przybyla B, Gurley C, Harvey JF, Bearden E, Kortebein P, Evans WJ, Sullivan DH, Peterson CA, Dennis RA (2006) Aging alters macrophage properties in human skeletal muscle both at rest and in response to acute resistance exercise. *Exp Gerontol* 41(3):320–327
55. Pyne DB, Baker MS, Fricker PA, McDonald WA, Telford RD, Weidemann MJ (1995) Effects of an intensive 12-wk training program by elite swimmers on neutrophil oxidative activity. *Med Sci Sports Exerc* 27:536–542
56. Sakamoto Y, Shouzoh U, Shimanuki H, Kasai T, Takato J, Ozaki H, Kawakami Y, Haga H (2005) Effects of low-intensity physical exercise on acute changes in resting saliva secretory IgA levels in the elderly. *Geriatr Gerontol Int* 5:202–206
57. Shephard RJ, Shek PN (1995) Cancer, immune function, and physical activity. *Can J Appl Physiol* 20(1):1–25
58. Shimizu K, Kimura F, Akimoto T, Akama T, Kuno S, Kono I (2007) Effect of free-living daily physical activity on salivary secretory IgA in elderly. *Med Sci Sports Exerc* 39(4):593–598
59. Shinkai EM, Rodrigues MF, Krause MS, Vianna DR, Almeida BS, Rossato JS, Oliveira LP Jr, Curi R, de Bittencourt PI Jr. (2007) Acute exercise stimulates macrophage function: possible role of NF-kappaB pathways. *Cell Biochem Funct* 25(1):63–73
60. Silveira EM, Rodrigues MF, Krause MS, Vianna DR, Almeida BS, Rossato JS, Oliveira LP Jr, Curi R, de Bittencourt PI Jr. (2007) Acute exercise stimulates macrophage function: possible role of NF-kappaB pathways. *Cell Biochem Funct* 25(1):63–73
61. Smith JA, Telford RD, Baker MS, Hapel AJ, Weidemann MJ (1992) Cytokine immunoreactivity in plasma does not change after moderate endurance exercise. *J Appl Physiol* 73:1396–1401
62. Smith JA, Gray AB, Pyne DB, Baker MS, Telford RD, Weidemann MJ (1996) Moderate exercise triggers both priming and activation of neutrophil subpopulations. *Am J Physiol* 270: R838–R845
63. Stout RD, Suttles J (2005) Immunosenescence and macrophage functional plasticity: dysregulation of macrophage function by age-associated microenvironmental changes. *Immunol Rev* 205:60–71
64. Straub RH, Miller LE, Schölmerich J, Zietz B (2000) Cytokines and hormones as possible links between endocrinosenescence and immunosenescence. *J Neuroimmunol* 109:10–15
65. Tsigos C, Papanicolaou DA, Defensor R, Mitsiadis CS, Kyrou L, Chrousos GP (1997) Dose effects of recombinant human interleukine-6 on pituitary hormone secretion and energy expenditure. *Neuroendocrinology* 66:54–62
66. Venyatraman JT, Fernandes G (1997) Exercise, immunity and aging. *Ageing* 9(1–2):42–56
67. Wenisch C, Patruta S, Daxbock F, Krause R, Horl W (2000) Effect of age on human neutrophil function. *J Leukoc Biol* 67:40–45
68. Yan H, Kuroiwa A, Tanaka H, Shindo M, Kiyonaga A, Nagayama A (2001) Effect of moderate exercise on immune senescence in men. *Eur J Appl Physiol* 86(2):105–111
69. Zanni F, Vescovini R, Biasini C, Fagnoni F, Zanlari L, Telera A, Di Pede P, Passeri G, Pedrazzoni M, Passeri M, Franceschi C, Sansoni P (2003) Marked increase with age of type 1 cytokines within memory and effector/cytotoxic CD8⁺T cells in humans: a contribution to understand the relationship between inflammation and immunosenescence. *Exp Gerontol* 38:981–987